

Axial Coordination of Carboxylate Activates the Non-heme Fe^{IV}=O Unit**

Jan-Uwe Rohde and Lawrence Que, Jr.*

High-valent oxoiron intermediates have been implicated in the dioxygen activation mechanisms of many mononuclear non-heme iron enzymes as the oxidizing species in metabolically important transformations.^[1] Recently, the first such species was directly observed in the catalytic cycle of TauD, a member of the superfamily of enzymes with a common 2-His-1-carboxylate facial triad motif that binds the active-site iron center.^[2–4] This short-lived intermediate has a high-spin iron(IV) center and the Fe–O bond is 1.62 Å long (determined by extended X-ray absorption fine structure spectroscopy (EXAFS)).^[5] Within the same time frame, we reported the first synthetic examples of mononuclear non-heme oxoiron(IV) complexes supported by tetradentate N4 (TMC and TPA) and pentadentate N5 (N4Py and Bn-TPEN) ligands.^[6–10] In the case of the TMC complex, [Fe^{IV}(O)(TMC)(NCMe)](OTf)₂ was sufficiently stable at –40 °C to be crystallized and characterized by X-ray diffraction.^[6] The synthetic complexes, in contrast to the TauD intermediate, have low-spin iron(IV) centers.^[6–9] The difference in spin states may arise from the nitrogen-rich nature of the iron coordination environment in the model complexes and the involvement of carboxylate ligands in TauD. This comparison prompted us to investigate whether a carboxylate ligand can be introduced into the iron coordination sphere of a synthetic oxoiron(IV) complex by ligand exchange and thus to assess how such a ligand substitution may affect the spectroscopic properties of the oxoiron(IV) center and its reactivity. We report herein on the reaction of [Fe^{IV}(O)(TMC)(NCMe)]²⁺ (**2**-NCMe) with NEt₄CF₃CO₂ and describe the properties of the new intermediate [Fe^{IV}(O)(TMC){OC(O)CF₃}]⁺ (**2**-O₂CCF₃).

The reaction of [Fe^{IV}(O)(TMC)(NCMe)]²⁺ (**2**-NCMe, λ_{max} = 824 nm, ε = 400 M^{–1} cm^{–1}) with 10 equivalents of NEt₄CF₃CO₂ in MeCN at –20 °C affords a new species designated **2**-O₂CCF₃ within 20 minutes, as indicated by the shift of the absorption maximum to 836 nm with a concomitant loss in intensity (ε = 250 M^{–1} cm^{–1}; Figure 1 a). Additional features are also observed to grow at around 940 and 990 nm. This conversion is marked by an isosbestic point, which suggests that **2**-O₂CCF₃ is produced directly from **2**-NCMe (Figure 1 a, Scheme 1). Complex **2**-O₂CCF₃ can also be generated by oxygen-atom transfer from PhIO to

[*] Dr. J.-U. Rohde, Prof. Dr. L. Que, Jr.
Department of Chemistry and
Center for Metals in Biocatalysis
University of Minnesota
207 Pleasant Street SE, Minneapolis, MN 55455 (USA)
Fax: (+1) 612-624-7029
E-mail: que@chem.umn.edu

[**] This research was supported by the National Institutes of Health (GM-33162). J.-U.R. acknowledges support by the Deutsche Forschungsgemeinschaft.

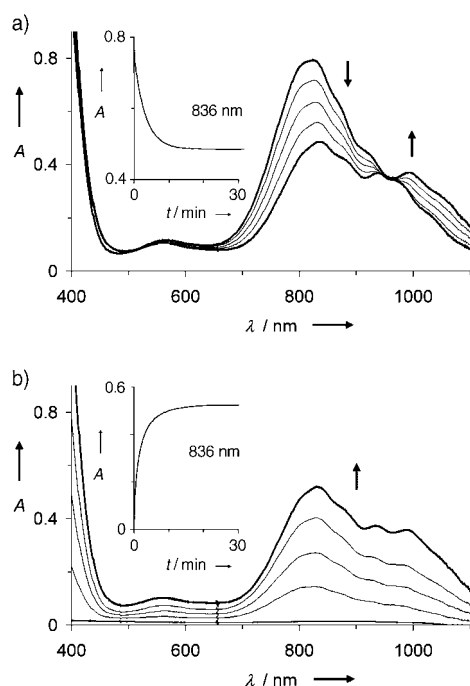
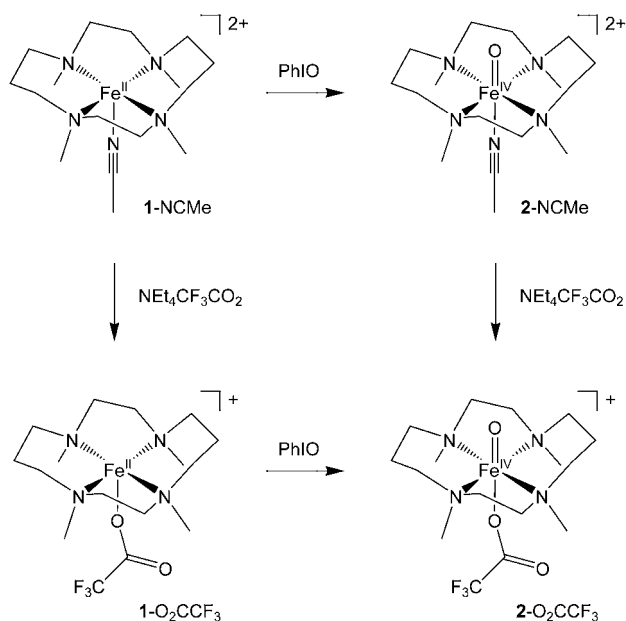


Figure 1. Generation of **2-O₂CCF₃** by a) addition of NEt₄CF₃CO₂ (10 equiv) to **2-NCMe**, and b) addition of PhIO (1 equiv) to **1-O₂CCF₃** as monitored by UV/Vis/NIR spectroscopy. Reaction conditions: 2 mm iron complex in MeCN at -20°C . Insets: Corresponding time courses of the reactions monitored at 836 nm.



Scheme 1. Ligand-substitution and oxygen-atom-transfer reactions of Fe(TMC) complexes.

[Fe^{II}(TMC){OC(O)CF₃}]⁺ (**1-O₂CCF₃**; Figure 1 b). Complex **1-O₂CCF₃** is formed in situ by treatment of [Fe^{II}(TMC)(NCMe)]²⁺ (**1-NCMe**) with 1 equivalent of NEt₄CF₃CO₂ as indicated by the shift of the very weak ligand-field band at $\lambda \approx 780 \text{ nm}$ to $\lambda \approx 890 \text{ nm}$. Thus, **2-O₂CCF₃** can be formed from

1-NCMe by two pathways, both involving ligand exchange and oxygen-atom transfer but in different order (Scheme 1).

Direct evidence for the binding of trifluoroacetate to the metal centers in **1-O₂CCF₃** and **2-O₂CCF₃** is provided by ¹⁹F NMR spectroscopy. In CD₂Cl₂, the starting material [Fe^{II}(TMC)(OTf)]OTf has two equally intense ¹⁹F resonance signals at $\delta = 15.6 \text{ ppm}$ and -85.2 ppm (-20°C); the signal at $\delta = -85.2 \text{ ppm}$ corresponds to free triflate ion in CD₂Cl₂ indicating that the signal at $\delta = 15.6 \text{ ppm}$ arises from triflate that is coordinated to the paramagnetic high-spin iron(II) center, [Fe^{II}(TMC)(OTf)]OTf (Figure 2 a). In CD₃CN, only

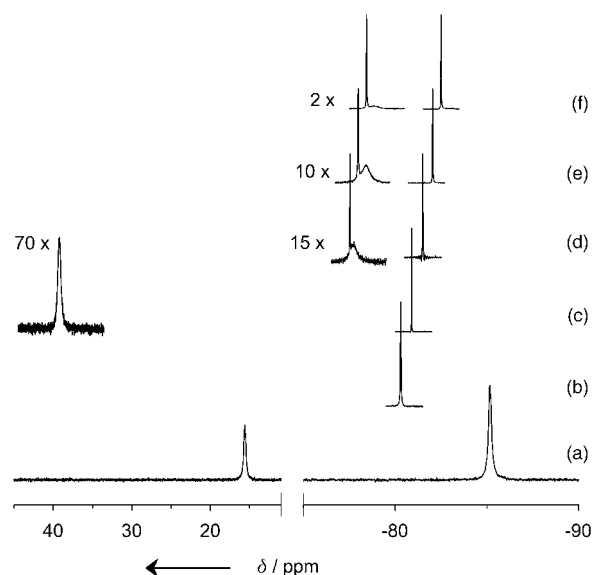


Figure 2. ¹⁹F NMR spectra: a) [Fe^{II}(TMC)(OTf)]OTf in CD₂Cl₂ (**1-OTf**) and b) in CD₃CN (**1-NCCD₃**), -20°C ; c) equimolar amounts of [Fe^{II}(TMC)(OTf)]OTf and NEt₄CF₃CO₂ in CD₃CN (**1-O₂CCF₃**), -20°C ; d) [Fe^{IV}(O)(TMC){OC(O)CF₃}]OTf in CD₃CN (**2-O₂CCF₃**) at -20°C and e) at -40°C ; and f) after addition of 1 equivalent of NEt₄CF₃CO₂ to **2-O₂CCF₃** in CD₃CN, -40°C .

one peak at $\delta = -79.9 \text{ ppm}$ is observed, which is attributed to the free triflate counterions (Figure 2 b). Addition of 1 equivalent of NEt₄CF₃CO₂ to this solution results in the appearance of a resonance at $\delta = 40.3 \text{ ppm}$, which corresponds to CF₃CO₂[−] ligated to high-spin Fe^{II}, with half the intensity of the peak at $\delta = -80.0 \text{ ppm}$ arising from the free triflate counterions (Figure 2 c). Addition of a second equivalent of NEt₄CF₃CO₂ affords a new peak at $\delta = -76.0 \text{ ppm}$ arising from free CF₃CO₂[−] ions (not shown), indicating that the iron center can only bind one trifluoroacetate. Upon addition of PhIO to **1-O₂CCF₃** at -20°C , the resonance of bound CF₃CO₂[−] disappeared and two new features could be observed close to $\delta = -76 \text{ ppm}$ (Figure 2 d). The broader peak is assigned to the CF₃CO₂[−] ligated to the paramagnetic Fe^{IV} center ($\delta = -76.3 \text{ ppm}$, 80 %), while the sharper peak corresponds to residual free CF₃CO₂[−] ($\delta = -76.1 \text{ ppm}$, 20 %). In support, addition of another equivalent of NEt₄CF₃CO₂ results in an increased intensity for the sharper peak (Figure 2 f). Thus trifluoroacetate can displace the axial MeCN ligand in **2-NCMe** to form **2-O₂CCF₃**.

Interestingly, the binding of trifluoroacetate affects the stability and reactivity of the oxoiron(IV) unit (Table 1). As previously reported, **2**-NCMe is unexpectedly rather stable and quite unreactive, persisting for at least a month at -40°C ,

Table 1: Properties of **2**-NCMe and **2**-O₂CCF₃.

	2 -NCMe	2 -O ₂ CCF ₃
λ_{max} [nm] (ϵ [$\text{M}^{-1}\text{cm}^{-1}$])	824 (400)	836 (250)
$k_{\text{subst.}}$ [s^{-1}] (-20°C) ^[a]	—	0.0053 (2)
for 2 -NCMe \rightarrow 2 -O ₂ CCF ₃		
k_{decay} [s^{-1}] (25°C) ^[b]	$1.8(2) \times 10^{-5}$	$2.2(4) \times 10^{-4}$
$t_{1/2}$ (25°C)		
in the absence of substrates ^[b]	10 h	1 h
+ 1 PPh ₃ ^[c]	≈ 20 s	≈ 20 s
+ 10 C ₁₄ H ₁₂ ^[d]	1 min	< 5 s

Reaction conditions: [a] Addition of NEt₄CF₃CO₂ (10 equiv) to 2 mM **2**-NCMe in MeCN at -20°C . [b] Addition of NEt₄CF₃CO₂ (1 equiv) to 2 mM **2**-NCMe in MeCN at 25°C . [c] Addition of PPh₃ (1 equiv) to 2 mM [Fe^{IV}(O)(TMC)(L)]²⁺ (**2**-L) in MeCN at 25°C . [d] Addition of dihydroanthracene (10 equiv) to 2 mM **2**-L in MeCN at 25°C .

thereby allowing crystals to be obtained.^[6] PPh₃ is the only substrate reported to date to be oxidized at -40°C by **2**-NCMe^[7] and the oxo transfer to one equivalent of substrate occurs over the course of an hour. Raising the temperature to 25°C shortens its half-life to 10 h and to 20 s in the presence of one equivalent PPh₃. Furthermore, **2**-NCMe reacts with 10 equivalents of dihydroanthracene to form anthracene (half-life of 1 min), so **2**-NCMe appears capable of hydrogen-atom abstraction from weak C–H bonds at room temperature. The yield of anthracene is 45 %, the same as that observed for the much more reactive [Fe^{IV}(O)(N4Py)]²⁺,^[11] suggesting that both oxoiron(IV) units act as one-electron oxidants in this transformation.

In contrast, **2**-O₂CCF₃ has a half-life at 25°C an order of magnitude shorter ($t_{1/2} \approx 1$ h; Table 1) than **2**-NCMe. Furthermore, **2**-O₂CCF₃ reacts much faster than **2**-NCMe with 10 equivalents of dihydroanthracene ($t_{1/2} \approx 5$ s) to produce approximately 0.4 equivalents of anthracene. However, the reaction of **2**-O₂CCF₃ with one equivalent of PPh₃ to afford OPPh₃ and regenerate **1**-O₂CCF₃, as monitored by ³¹P and ¹⁹F NMR spectroscopy, occurs on the same time scale as that of **2**-NCMe. So axial coordination of carboxylate to the Fe^{IV}=O unit appears to convert **2**-NCMe into a more reactive oxidant for one-electron oxidations, but not for oxygen atom transfer onto a phosphorus, formally a two-electron process.

The observed lower thermal stability and greater reactivity of **2**-O₂CCF₃ towards dihydroanthracene may at first glance appear counterintuitive, as the replacement of neutral MeCN with a monoanionic [CF₃CO₂][−] ligand might be expected to stabilize the iron(IV) oxidation state by reducing its high effective charge. Indeed Collins has used this principle in his design of tetraanionic macrocyclic tetraamidate ligands that led to the isolation and crystallization of novel iron(IV) complexes.^[12] On the other hand, Borovik and co-workers have reported the first example of an iron(III) complex with a terminal oxo ligand, [Fe^{III}(O)(L)]^{2−}, where L is a tripodal trianionic tris(ureaylato)amine ligand, which is derived from

the reaction of O₂ with the iron(II) precursor. They have invoked the involvement of an oxoiron(IV) species that is too reactive to be detected as the precursor to the oxoiron(III) complex.^[13,14] Also, Wieghardt and co-workers have observed the formation of a rather unstable oxoiron(IV) complex containing a cyclam ligand with a carboxymethyl tether from the reaction of its iron(III) precursor with O₃.^[15,16] Our results, together with the latter two observations above, suggest that introducing anionic ligands can in fact destabilize the oxoiron(IV) center. In the case of **2**, replacing the nitrile with a carboxylate results in the weakening of the ligand field, as indicated by the red shift observed for the near-IR d–d transitions associated with the $S=1$ oxoiron(IV) unit of **2**-O₂CCF₃ relative to those of **2**-NCMe,^[17] and a consequent decrease in the low-spin/high-spin energy gap. The higher reactivity observed for **2**-O₂CCF₃ may thus be associated with enhanced access to a more reactive high-spin $S=2$ reaction surface, although the current limited information available on oxoiron(IV) centers indicates that the strength of the Fe=O bond is comparable for low-spin and high-spin complexes.^[4–6,9,17] Nevertheless, there has been considerable discussion on the spin-state-dependent reactivity of oxoiron(IV) species in heme systems to rationalize some of the conflicting observations that have been made for the nature of the active oxidant.^[18,19] Similar considerations may apply to non-heme oxoiron(IV) oxidations.

In summary, we have introduced a carboxylate ligand *trans* to an oxo group in an iron(IV) complex and, in so doing, have enhanced the reactivity of the $S=1$ oxoiron(IV) unit in **2**.^[20] We hypothesize that the higher reactivity may be due to a weakening of the ligand field that provides greater access to a more reactive $S=2$ surface. We note that such $S=2$ iron(IV) centers are utilized by non-heme iron enzymes, such as methane monooxygenase, ribonucleotide reductase, and TauD, to carry out their respective oxidative reactions, and the common feature among these enzymes is the presence of histidine and carboxylate ligands.^[1,21,22]

Experimental Section

Physical Methods: UV/Vis spectra were recorded on an HP8453 A diode array spectrometer with samples maintained at low temperature by using a cryostat from Unisoku Scientific Instruments, Japan. ¹⁹F and ³¹P NMR spectra were recorded on a Varian Inova VXR-300 or Varian Inova VI-300 spectrometer at ambient temperature, unless noted otherwise. Chemical shifts (ppm) are reported referenced to an external standard, CFCl₃ ($\delta=0$ ppm) for ¹⁹F NMR spectra and H₃PO₄ (85 %, 0 ppm) for ³¹P NMR spectra. GC analyses were performed on a Perkin-Elmer Sigma 3 gas chromatograph (AT-1701 column, 30 m) with flame-ionization detection. The products were identified by comparing their retention times to those of authentic samples.

Generation of [Fe(O)(TMC){OC(O)CF₃}]OTf (**2**-O₂CCF₃(OTf)) via [Fe(O)(TMC)(NCMe)](OTf)₂ (**2**-NCMe(OTf)₂). A 2 mM solution of [Fe^{II}(TMC)(OTf)]OTf^[6] in MeCN (2.5 mL, 0.005 mmol Fe) in a 1-cm UV/Vis cuvette was pre-cooled to -20°C . (Note: Dissolution of [Fe^{II}(TMC)(OTf)]OTf in MeCN readily afforded [Fe(TMC)(NCMe)](OTf)₂ (**1**-NCMe(OTf)₂), as indicated by the ¹⁹F resonance of the free triflate ion in its NMR spectrum.) Addition of PhIO^[23] (1 equiv, 0.005 mmol) in MeOH (0.05 mL) afforded **2**-NCMe(OTf)₂. Subsequent addition of NEt₄CF₃CO₂ (10 equiv, 0.050 mmol) in MeCN (0.05 mL) afforded **2**-O₂CCF₃(OTf). ¹⁹F NMR for **1**-OTf(OTf)

(CD₂Cl₂, 282.4 MHz, –20 °C): δ = 15.6 (s, [Fe(TMC){OS(O)₂CF₃}]⁺) and –85.2 ppm (s, CF₃SO₃[–]); **1**-NCCD₃(OTf)₂ (CD₃CN, 282.4 MHz, –20 °C): δ = –79.9 ppm (s, CF₃SO₃[–]). ¹⁹F NMR for **2**-O₂CCF₃(OTf) + NEt₄OTf (CD₃CN, 282.4 MHz, –20 °C): δ = –76.1 (s, CF₃CO₂[–]), –76.3 (s, [Fe(O)(TMC){OC(O)CF₃}]⁺), and –80.0 ppm (s, CF₃SO₃[–]). ¹⁹F NMR for NEt₄CF₃CO₂ (CD₂Cl₂, 282.4 MHz): δ = –75.9 ppm (s, CF₃CO₂[–]); NEt₄CF₃CO₂ (CD₃CN, 282.4 MHz): δ = –75.8 ppm (s, CF₃CO₂[–]).

Generation of **2**-O₂CCF₃(OTf) via [Fe(TMC){OC(O)CF₃}]OTf (**1**-O₂CCF₃(OTf)). A 2 mm solution of [Fe^{II}(TMC)(OTf)]OTf in MeCN (2.5 mL, 0.005 mmol Fe) in a 1-cm UV/Vis cuvette was converted into **1**-O₂CCF₃(OTf) by addition of NEt₄CF₃CO₂ (1 equiv, 0.005 mmol) in MeCN (0.05 mL). This solution was then cooled to –20 °C. **2**-O₂CCF₃(OTf) was generated by addition of PhIO (1 equiv, 0.005 mmol) in MeOH (0.05 mL). ¹⁹F NMR for **1**-O₂CCF₃(OTf) + NEt₄OTf (CD₃CN, 282.4 MHz, –20 °C): δ = 40.3 (s, [Fe(TMC){OC(O)CF₃}]⁺) and –80.0 ppm (s, CF₃SO₃[–]).

Reaction of **2**-L with PPh₃. A 2 mm solution of **2**-NCMe(OTf)₂ or **2**-O₂CCF₃(OTf) in MeCN (2 mL), generated from **1**-NCMe(OTf)₂ or **1**-O₂CCF₃(OTf) and PhIO (1 equiv) at 25 °C, was treated with PPh₃ (1 equiv, 0.004 mmol) in a solvent mixture of MeCN and CH₂Cl₂ (1:1; 0.04 mL). Products were analyzed by ³¹P NMR (CD₃CN, 121.4 MHz): δ = 27.9 ppm (s, OPPh₃), and ¹⁹F NMR (CD₃CN, 282.4 MHz): δ = 21.4 (s, [Fe(TMC){OC(O)CF₃}]⁺) and –80.0 ppm (s, CF₃SO₃[–]).

Reaction of **2**-L with dihydroanthracene: A 2 mm solution of **2**-NCMe(OTf)₂ or **2**-O₂CCF₃(OTf) in MeCN (2 mL), generated by addition of PhIO (1 equiv) to **1**-NCMe(OTf)₂ or **1**-O₂CCF₃(OTf) at 25 °C, was treated with dihydroanthracene (10 equiv, 0.040 mmol; recrystallized from EtOH)^[24] in a solvent mixture of MeCN and CH₂Cl₂ (1:1; 0.08 mL). Products were analyzed by GC: Reaction with **2**-NCMe(OTf)₂ yielded 0.0018 mmol anthracene (45% based on equivalents of Fe complex) and reaction with **2**-O₂CCF₃(OTf) gave 0.0016 mmol anthracene (40%).

Received: November 16, 2004

Published online: February 28, 2005

Keywords: bioinorganic chemistry · carboxylate ligands · high-valent compounds · iron · oxo ligands

sulfonate (or triflate) anion, TMC = 1,4,8,11-tetramethyl-1,4,8,11-tetraazacyclotetradecane (or tetra(*N*-methyl)cyclam), TPA = *N,N,N*-tris(2-pyridylmethyl)amine.

- [1] M. Costas, M. P. Mehn, M. P. Jensen, L. Que, Jr., *Chem. Rev.* **2004**, *104*, 939.
- [2] J. C. Price, E. W. Barr, B. Tirupati, J. M. Bollinger, Jr., C. Krebs, *Biochemistry* **2003**, *42*, 7497.
- [3] J. C. Price, E. W. Barr, T. E. Glass, C. Krebs, J. M. Bollinger, Jr., *J. Am. Chem. Soc.* **2003**, *125*, 13008.
- [4] D. A. Proshlyakov, T. F. Henshaw, G. R. Monterosso, M. J. Ryle, R. P. Hausinger, *J. Am. Chem. Soc.* **2004**, *126*, 1022.
- [5] P. J. Riggs-Gelasco, J. C. Price, R. B. Guyer, J. H. Brehm, E. W. Barr, J. M. Bollinger, Jr., C. Krebs, *J. Am. Chem. Soc.* **2004**, *126*, 8108.
- [6] J.-U. Rohde, J.-H. In, M. H. Lim, W. W. Brennessel, M. R. Bukowski, A. Stubna, E. Münck, W. Nam, L. Que, Jr., *Science* **2003**, *299*, 1037.
- [7] M. H. Lim, J.-U. Rohde, A. Stubna, M. R. Bukowski, M. Costas, R. Y. N. Ho, E. Münck, W. Nam, L. Que, Jr., *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 3665.
- [8] J. Kaizer, E. J. Klinker, N. Y. Oh, J.-U. Rohde, W. J. Song, A. Stubna, J. Kim, E. Münck, W. Nam, L. Que, Jr., *J. Am. Chem. Soc.* **2004**, *126*, 472.
- [9] J.-U. Rohde, S. Torelli, X. Shan, M. H. Lim, E. J. Klinker, J. Kaizer, K. Chen, W. Nam, L. Que, Jr., *J. Am. Chem. Soc.* **2004**, *126*, 16750.
- [10] Abbreviations used: Bn-TPEN = *N*-benzyl-*N,N',N'*-tris(2-pyridylmethyl)-1,2-diaminoethane, N4Py = *N,N*-bis(2-pyridylmethyl)-*N*-bis(2-pyridyl)methylamine, OTf = trifluoromethyl-
- [11] E. J. Klinker, L. Que, Jr., University of Minnesota, unpublished results.
- [12] T. J. Collins, *Acc. Chem. Res.* **1994**, *27*, 279.
- [13] C. E. MacBeth, A. P. Golombek, V. G. Young, Jr., C. Yang, K. Kucera, M. P. Hendrich, A. S. Borovik, *Science* **2000**, *289*, 938.
- [14] C. E. MacBeth, R. Gupta, K. R. Mitchell-Koch, V. G. Young, Jr., G. H. Lushington, W. H. Thompson, M. P. Hendrich, A. S. Borovik, *J. Am. Chem. Soc.* **2004**, *126*, 2556.
- [15] C. A. Grapperhaus, B. Mienert, E. Bill, T. Weyhermüller, K. Wieghardt, *Inorg. Chem.* **2000**, *39*, 5306.
- [16] Wieghardt's putative oxoiron(IV) complex with a pentadentate ligand consisting of the macrocyclic cyclam ligand with a pendent acetate from one of the nitrogen atoms is a rather unstable species that was generated from the reaction of [Fe^{III}(cyclam-acetate)(OTf)]⁺ and O₃ at –80 °C in less than 25% yield and thus difficult to characterize.^[15] The proposed oxoiron(IV) formulation was made on the basis of a Mössbauer analysis, a hypothesis that was subsequently supported by the synthesis and characterization of more stable oxoiron(IV) complexes.^[6–9] The instability of this complex might be ascribed to the presumed ligation of the pendent carboxylate, consistent with the results reported herein, but might also derive from the presence of secondary amine ligands (versus tertiary amine ligands in **2**-NCMe and **2**-O₂CCF₃) that are more susceptible to ligand oxidation.^[25] The lack of more detailed structural information on this interesting complex however makes it difficult to draw firm conclusions about its properties.
- [17] A. Decker, J.-U. Rohde, L. Que, Jr., E. I. Solomon, *J. Am. Chem. Soc.* **2004**, *126*, 5378.
- [18] J. C. Schoeneboom, S. Cohen, H. Lin, S. Shaik, W. Thiel, *J. Am. Chem. Soc.* **2004**, *126*, 4017.
- [19] S. Shaik, S. P. de Visser, D. Kumar, *J. Biol. Inorg. Chem.* **2004**, *9*, 661.
- [20] Gross and co-workers have also reported that coordination of an axial carboxylate to an oxoiron(IV) porphyrin cation radical complexes similarly enhances its reactivity: Z. Gross, S. Nimri, C. M. Barzilay, L. Simkhovich, *J. Biol. Inorg. Chem.* **1997**, *2*, 492.
- [21] B. J. Wallar, J. D. Lipscomb, *Chem. Rev.* **1996**, *96*, 2625.
- [22] M. Merks, D. A. Kopp, M. H. Sazinsky, J. L. Blazyk, J. Müller, S. J. Lippard, *Angew. Chem. Int. Ed.* **2001**, *40*, 2782.
- [23] H. Saltzman, J. G. Sharefkin, *Organic Syntheses*, Coll. Vol. V, Wiley, New York, **1973**, p. 658.
- [24] W. L. F. Armarego, D. D. Perrin, *Purification of Laboratory Chemicals*, Butterworth-Heinemann, Oxford, **1997**.
- [25] D. H. Busch, N. W. Alcock, *Chem. Rev.* **1994**, *94*, 585.